The single breath transfer factor ($T_{L,CO}$) and the transfer coefficient (K_{CO}): a window onto the pulmonary microcirculation

J.M.B. Hughes

Division of Respiratory Medicine, National Heart and Lung Institute, Imperial College School of Medicine, Hammersmith Hospital Campus, London, UK

Summary

Correspondence

JMB Hughes, 4 Cedars Road, London SW13 0HP, UK E-mail: mike.hughes@ic.ac.uk

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The transfer factor, TLCO (with the transfer coefficient, KCO, also known as the transfer factor per unit alveolar volume, [TL/VA], is one of the most useful clinical tests of pulmonary function, the only one which specifically focuses on pulmonary microcirculation. It was originally devised in 1909 as a physiological tool to assess the diffusive capacity of the lung as a gas exchanger. It was subsequently developed as a clinical tool, but cumbersome analytical techniques delayed its introduction into clinical medicine until 1950s. The physiology of the carbon monoxide transfer factor (also called the diffusing capacity DLCO) is based on the Roughton-Forster equation which partitions DL,CO, a conductance, into membrane (DM) and red cell (θVc) diffusion conductances. Recent work (1987–2001) suggests that 70–80% of the resistance to CO (and O₂) diffusion may reside in the red cell fraction. The clinical implication is that TL,CO and KCO are 'windows' onto the pulmonary microcirculation. As regards reference values for clinical use, TL, co depends on age, height and gender. Kco, which is actually a rate constant, is independent of gender, and is affected principally by age. A schema is presented for the clinical interpretation of TL, co. As TL, co is derived from the product of Kco and the accessible alveolar volume (V_A), examination of these two components (Kco and V_A) will usually suggest a specific pathophysiological mechanism as the explanation for a reduction in TL,CO.

Introduction

The transfer factor of the lung for carbon monoxide (TL.CO), also known as the diffusing capacity (DL.CO), has become one of the key tests of pulmonary function. TL.CO measures the potential of the lung for gas exchange. For example, a patient with interstitial lung disease might have a low TL.CO (DL.CO), (say <50% predicted normal), but could still have a normal arterial PO₂ (PaO₂) at rest; but on exercise, PaO₂ will fall, often severely, because the lung has insufficient gas exchanging surface area to meet the additional oxygen demand. As will be seen later, the particular surface area which is crucial is that of the microvascular bed, in particular the number of capillaries.

History of the TL,co (DL,co) measurement

Most of our current pulmonary function tests were introduced in the 1950s. DL,CO, as it was called then, has a much longer history. It was devised originally by Krogh & Krogh (1909), in Denmark, by August Krogh and Marie (his wife), as a physiological tool to test the notion, long since abandoned, that the lung, like the swim

bladder of deep-sea fish, could secrete oxygen against the normal pressure gradient exerted by the inspired air. Subsequently, DL,CO was introduced as a clinical test by Krogh (1915), but the measurement never caught on because methods of measuring carbon monoxide (CO) - by combusting the gas with oxygen to produce CO₂ - were cumbersome. It was not until after the Second World War (1939-45), following the invention in Germany of the infra-red technique for CO detection that Marie Krogh's original measurements were repeated and refined for clinical use; the principle modification, suggested by W.S. Fowler (Forster et al., 1954a), was the addition of an inert gas (helium) to the CO–air mixture, so that CO_0 could be calculated (see Fig. 1) rather than measured. In M. Krogh's (1915) original technique, CO₀ was obtained from an initial expiration from total lung capacity (TLC) to mid-lung volume; after a breath hold of 6-8 s, a second expiration was made and CO_t sampled.

The diffusing capacity for oxygen

Lilienthal et al. (1946) published a landmark paper describing a method for measuring the oxygen diffusing capacity (DL,O_2) ,

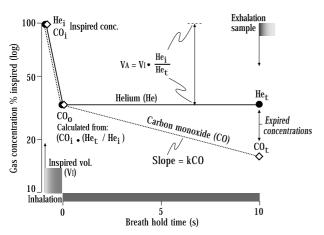


Figure 1 Concentrations of the test gases (carbon monoxide and helium) plotted against breath hold time for the single breath $T_{L,CO}$ manoeuvre illustrating the origin and calculation of the two components (Kco and VA) from which the $T_{L,CO}$ is derived. V_A = alveolar volume at which the Kco was measured.

for which DL,CO was after all only a substitute. It could be argued that this paper ushered in a new era in gas exchange research. The ingenuity of Lilienthal's method lay in the differentiation of end-capillary (Pc'O₂) from arterial (PaO₂) oxygen tension; this was achieved by measuring alveolar and arterial PO₂ on exercise breathing air, followed by a 10-12% O₂ hypoxic mixture. They were delighted when the steady state DLO₂ values were similar to Krogh's (1915) single breath DL,CO measurements, also made on exercise.

Steady state DL, co

With the revival of interest in diffusion measurements, several groups (Bates, 1952; Filley et al., 1954; Bates et al., 1955) thought it more appropriate to measure DL,CO by a steady state technique (DL,CO ss), similar to that for DLO₂. The measurement of DL,CO ss was a much easier proposition than DLO2. There was no need to repeat the measurement under hypoxic conditions, $P\bar{c}$ (the mean capillary tension) for CO could be ignored (haemoglobin being considered an infinite sink for CO), and DL,CO ss computed as \dot{V}_{CO} /PACO, where \dot{V}_{CO} is CO uptake in ml min⁻¹, and PA is the alveolar tension. [note: $D_LO_2 = (\dot{V}O_2 / I)$ $(PAO_2 - P\bar{c}O_2)$]. The measurement of DL, co ss was straightforward, but the assessment of PACO by direct sampling was problematic in the presence of a sloping alveolar plateau of CO concentration on expiration, as was frequently the case in respiratory disease. The solution of Filley et al. (1954) was to calculate an 'ideal' PACO, as for the alveolar-arterial gradient for oxygen, although this involved taking an arterial sample for PaCO2. A few simultaneous comparisons, mostly in normal subjects, were made on exercise of DL,CO ss and DLO2 and there was reasonable concordance (Forster et al., 1955; Shepard et al., 1958). DL,CO sb exceeded DL,CO ss at rest by about 33% (Marshall, 1958), but the two measurements converged on exercise.

Steady state or single breath DL,co?

In the late 1950s, Ogilvie et al. (1957) published their 'standardized technique' for the single breath DL,CO, called DL,CO sb, incorporating Fowler's helium modification (Forster et al. (1954a); see earlier]. In the 1960s, DL,CO ss or DL,CO sb were being introduced in the pulmonary function laboratories of most University Departments. By the 1970s, the concept and clinical usefulness of DL,CO (now called TL,CO throughout Europe) was widely accepted and it was the single breath technique which became the method of choice. The reasons were pragmatic. DL,CO sb did not require an arterial sample, nor meticulous timing of alveolar samples. It was more acceptable to patients and staff. In 1965, an automated apparatus for measuring DL, co sb came onto the market (Meade et al., 1965). It was a success and other manufacturers followed suit. Once DL, co became a 'black box' test, any inhibitions clinicians might have had about setting it up were quickly dispelled!

Physiology of DL, co (TL, co)

The Kroghs believed that CO uptake from alveolar gas occurred by passive diffusion across the alveolar-capillary membranes, driven by the PA-Pc gradient for CO. Krogh & Krogh (1909) converted DL,CO to DL,O2 by multiplying by 1.23 which is the ratio of the tissue diffusivities of O₂ and CO. Following Christian Bohr (1909) who was August Krogh's teacher, they reasoned that PcCO would be negligible because of the high affinity of CO for Hb. As we have seen, this assumption greatly simplifies the measurement. Like many assumptions, it proved to be only partially correct (see later)! Nevertheless, until the 1940s, resistance to gas exchange was considered to be 'diffusive', i.e. proportional to the thickness/ surface area ratio of the intervening membranes and tissues. For pulmonary gas exchange, the final step for oxygen, the combination with haemoglobin to form HbO2, was thought to be practically instantaneous (just a few milliseconds). Roughton (1932), on the other hand, [from his work with Hartridge (Hartridge & Roughton, 1923)], noted that the reaction velocity for O_2 or CO was seven to 10 times slower in intact red cells than in haemoglobin solutions, and he concluded that diffusion resistance of the red cell membrane and the interior of the cell was responsible.

In the 1950s, R.E. Forster was asked by Julius Comroe (Head of Physiology at the University of Pennsylvania) to repeat Krogh's (1915) measurements with the aim of developing a clinically useful test (Comroe, 1975). Forster became (fortunately for us!) sidetracked by his desire to understand exactly what was being measured by $D_{L,CO}$ (Forster et al., 1954b). His research, with several collaborators including Roughton (a frequent visitor to Philadelphia from his laboratory in Cambridge), culminated in the publication of a famous article (Roughton & Forster, 1957) containing the equation:

$$\frac{1}{D_{\rm L}} = \frac{1}{D_{\rm M}} + \frac{1}{\theta V_{\rm C}} \tag{1}$$

where DM is the diffusing capacity of the membranes separating the alveolar gas from the red cell itself; θ is the rate of reaction of CO with red cells (sometimes called the diffusing capacity per ml of blood), and Vc is the microvascular (capillary) blood volume in contact with the inhaled CO. Kruhoffer (1954), who worked in the same Department in Copenhagen as did the Kroghs, had published a similar equation 3 years earlier, but without giving a formal proof, and with erroneous values for θ .

The Roughton–Forster equation is the key to understanding what the DLCO is measuring. DLCO is a conductance (ml min⁻¹ mmHg⁻¹), and the reciprocals in equation (1) are resistances. $1/\theta$ VC is the oxygen-dependent part of the resistance, located in the pulmonary capillaries, and 1/DM is what remains when 1/DL is back-extrapolated to zero PO₂. [1/DL - 1/DM]/[1/DL] is the 'red cell' fraction of the total diffusion resistance. Roughton & Forster (1957) also showed, from measurements of DLCO at two or more alveolar PO₂ levels (ideally 150–200 and 500–600 mmHg) and knowing θ CO at the appropriate PAO₂, that DM and VC could be calculated by a simple graphical method.

The membrane diffusing capacity

The DM is determined by the tissue diffusivity [solubility/ (mol. wt)⁻²] of O_2 or CO in lung tissue and the surface area/ thickness ratio of the epithelial, interstitial, endothelial and plasma barriers. In physiological terms, DM is a function of the expansion of the lung. With a doubling of the gas volume of the lung (from 50 to 100% TLC) DM increases by about 75%, but DL co by only 25% (Vc does not change) (Stam et al., 1991). Increases of DM as the lung expands are caused by a mixture of airspace spherical expansion and unfolding of the surface. Unfortunately, the measurement of DM is not independent of Vc, because intracapillary Hb must be present for DM to be detected. Also, changes in the dimensions of the Vc component will alter DM.

The diffusing capacity of blood

In vivo, θ CO is inversely proportional to PO₂. The original in vitro measurements of Roughton & Forster (1957) were carried out at pH 7.8–8.0. More recent estimates (Forster, 1987) at physiological pH have resulted in significantly lower $1/\theta$ values at high PO₂s. If, in the Roughton–Forster equation, the 1987 $1/\theta$ are substituted for the 1957 ones, DM increases 2.5–3.5 times, and Vc decreases by 33%. Nearly all published DM and Vc values have used the 1957 θ CO values, a notable exception being Borland & Cox (1991) and Borland et al. (2001).

Pulmonary capillary volume

As measured by the Roughton–Forster technique, capillary volumes (with 1957 $1/\theta$) are in the range 80–100 ml

(females) and 100–120 ml (males) at rest, and 125–210 ml (males and females) on exercise (Hsia et al., 1995). These values would be about 33% lower if the 1987 $1/\theta$ were to be used. Morphometric values for Vc at rest, from post-mortem lungs, are about 200 ml (ranging from 120–280 ml depending on body weight) (Gehr et al., 1978). Because of the Fåhreus–Lindqvist effect, whereby red cells accelerate relative to mean plasma flow in their passage through the capillary bed, capillary haematocrit (Hct) is less than that in larger vessels. In fact, pulmonary capillary Hct is about 67% of large vessel Hct (Brudin et al., 1986). This effect does not change Vc estimates because there is an equal and opposite reduction in θ CO.

Diffusing capacity for nitric oxide

In the last 15 years, the alveolar uptake of nitric oxide (NO) has been studied (Guenard et al., 1987; Borland & Higenbottam, 1989). The theory and technique for estimating DL,NO is identical to that for DL,CO. DL,NO is four to five times greater than DL,CO. The reason is that θ NO is nearly seven times larger than θ CO (Carlsen & COmroe, 1958). From simultaneous inhalation of NO and CO, DL,NO and DL,CO can be calculated, and the Roughton–Forster equation solved on the basis of two values of $1/\theta$ ($1/\theta$ NO and $1/\theta$ CO), instead of two values of $1/\theta$ co at different PAO₂ s (Borland & COX, 1991; Borland et al., 2001). This has the effect of increasing DM,CO by three to four times, and reducing the VC/DM ratio from about 2 to 0.35.

Red cell resistance fraction

As mentioned earlier, the general view, before the Roughton & Forster (1957) paper, was that DL,CO~DM with no resistance to diffusion attributable to the reaction of CO with haemoglobin. When DL,CO was partitioned into DM and VC using the original 1957 θ co values, the red cell resistance fraction varied from 32-56% (Forster, 1957). With the morphometric analysis, the red cell resistance fraction is in the range 50-80% (Gehr et al., 1978). Using the 1987 θ co values and Borland & Cox (1991) and Borland et al.'s (2001) $\theta_{NO}-\theta_{CO}$ technique, this fraction has risen to 80%, i.e. most of the diffusion resistance is intracapillary. Thus, DL,CO measurements may be heavily weighted towards the numbers of red cells and/ or the number of capillary vessels. This new perception supports the views of clinicians who have maintained that DL,CO is a 'window on the pulmonary microcirculation'. Striking changes in DL, co in severe anaemia (\downarrow) [Rankin et al., 1961], on exercise (\uparrow) [Hsia et al., 1995], in intrapulmonary haemorrhage (\uparrow) [Ewan et al., 1976], and in pulmonary vasculitis (\downarrow) emphasize the pre-eminent role of the capillary bed.

The ratio $D_{L,O_2}/D_{L,CO}$ in hypoxia is about 1.2 (Meyer et al., 1981), the same as predicted (but for normoxia) by Krogh and Krogh in 1909. If the $D_{L,CO}$ is measured in normoxia, the D_{L,O_2} / $D_{L,CO}$ ratio is 1.7, reflecting a combination of O_2 /CO ratios for D_M (1.23) and θ [c. 2.0; Forster, 1987).

Perhaps surprisingly, partitioning the DL.CO into its DM and VC components (not difficult to do even in patients) has not proved useful clinically. There are two reasons: first, DL.CO is dominated by its θ VC component, and secondly, DM and VC measurements are coupled in the sense that VC must exist for DM to be measurable. The only examples of 'uncoupling' are congestive heart failure (Puri et al., 1995) (DM is reduced when VC is normal or high) and intrapulmonary haemorrhage (Ewan et al., 1976) (VC high, DM normal or reduced).

Clinical interpretation of TL, co and K co

Marie Krogh (1915) pointed out that the single breath TL,CO was the product of two separate measurements – the rate constant for CO removal from alveolar gas (which she called kco) and the alveolar volume (VA). This simple concept is the key, in our opinion (Hughes & Pride, 2001), to its clinical interpretation. kco is measured as the exponential decay in fractional concentration of CO over a period of breath-holding (BHT) — see Fig. 1:

$$kco = [log_e(CO_0/CO_t)]/BHT$$
(2)

where CO_0 and CO_t are the alveolar CO concentrations at the start and finish of BHT. The units of kco are s⁻¹ or min⁻¹.

The total CO transfer of the lungs is calculated as:

$$T_{L,CO} = [k_{CO} \times V_{A} \text{ stpd}] / [P_{B} - P_{H_{2}O}]$$
(3)

where P_B and P_{H_2O} are the barometric pressure and the water vapour pressure (at 37°C) which standardize for the driving pressure for CO uptake, i.e. the pressure of CO in the alveoli (PACO). The units of TL_{CO} are mmol min⁻¹ kPa⁻¹ (SI) and ml min⁻¹ mmHg⁻¹ (traditional).

In the original clinical description (Ogilvie *et al.*, 1957), and until 1965 when automated apparatus (and calculations) were introduced (Meade *et al.*, 1965), TLC was calculated independently from closed-circuit inert gas dilution (or body plethysmography) and used as 'VA'. In the absence of airflow obstruction, VA and TLC are approximately the same, but single breath VA may be considerably less than TLC when gas mixing is slow as in airflow obstruction (see later). Nowadays, the simultaneously measured single breath VA has replaced a separate measurement of TLC for logistic reasons; clinicians request TL, co more frequently than TLC.

It is our contention that the logical way to interpret TL,CO, in the clinical context, is in terms of its components (VA and kCO) from which it is derived. Unfortunately, the simplicity of this approach has been obscured by modern nomenclature. Today, Krogh's kCO is deployed in different units (although it is the same rate constant) as the carbon monoxide transfer coefficient (KCO), whose units of mmol min⁻¹ kPa⁻¹ L⁻¹ BTPS (in SI units) give misleadingly the appearance of being a ratio, an impression enhanced by its alternative terminology (TL/VA or DL/VA). In SI units, kCO [min⁻¹] converts to KCO (TL/VA) by dividing by 2.56, and in traditional units by dividing by 0.853.

Nomenclature

The Kroghs term for the DLCO was 'diffusion constant', but 'diffusing capacity' replaced it in the 1950s. John Cotes proposed (Cotes & Meade, 1963 'transfer factor' (TLCO) in recognition of the θ VC term in the Roughton–Forster equation. TLCO is in general use throughout Europe, although DLCO remains in use in North America. Krogh (1915) referred to kco as the 'permeability' factor. In modern parlance, the TL/VA or DL/VA is referred to as the 'transfer factor per unit lung volume', although 'transfer coefficient' is now the preferred term in Europe. Reflecting the original Krogh concept, the term Kco is replacing TL/VA or DL/VA.

Reference values

M. Krogh (1915) found that $D_{L,CO}$ was greater in men than in women, and Ogilvie *et al.* (1957) described the dependence of $D_{L,CO}$ on body surface area. Modern reference equations for TL,CO have height and age as coefficients, with separate regressions for men and women. The regression on height is determined by the V_A component of TL,CO (V_A in normal subjects being a surrogate for TLC). The regression on age is largely determined by the kco component.

Cotes & Hall (1970) pointed out that in young adults Kco was the same in both sexes, but declined with age at a faster rate in men than in women. For a man and a woman who started with the same KCO at age 25 years (say 1.8 in SI units), the male Kco at age 65 years would be 16% less than the woman's (from Cotes & Hall, 1970). Population studies of Kco, which have included both men and women, show a dependence on age with the coefficient being significantly slightly greater in men $(-0.023 \text{ years}^{-1} \text{ in males versus } -0.016$ years⁻¹ in females) (Hughes & Pride, in preparation). Most of the studies showed, in addition, a dependence on height, although there is no logical reason why a rate constant, which is what KCO ($\sim TL/VA$) actually represents, should be dependent on stature or gender. The dependence on height is intriguing. Gulsvik et al. (1992) have made the interesting suggestion that, in the seated position, and in taller people, the apices of the lungs may be more poorly perfused relative to the mid and lower zones for gravitational reasons; the resulting inhomogeneity in blood flow and blood volume would reduce the measured Kco for taller people.

In a review of the literature, we have found no significant gender difference for Kco at age 45 years, although a small (non-significant) difference emerges at age 65 years. The current EEC recommendations (Cotes et al., 1993) for reference values for Kco (\sim TL/VA) are based on [TL,co(predicted)] / TLC (predicted)]. The use of separate predictors, each with individual gender, height and (plus age for the TL,co) coefficients, for the Kco, which is actually a rate constant, seems illogical. Further investigation seems to be required.

Interpretation of TL, co and K co in lung disease

From eqn (3), $T_{L,CO} = k_{CO} \times V_{A}$. Therefore, a low $T_{L,CO}$ must be caused by a low Kco or a low VA or a combination of the two. It is also possible for Kco to be high (as a percentage of that expected at the predicted TLC). As a mechanical analogy, note that $FEV_1 = FEV_1 / VC \times VC$, and the explanation for a low FEV_1 must be either a low FEV_1/VC or a low VC or a combination (like Kco, FEV_1/VC may be high). Note that the single breath TL,CO is performed at full inflation, close to TLC, in the seated position, and at rest. In the absence of airflow obstruction with impaired gas mixing, the single breath estimate of VA should approach that of TLC minus the anatomic dead space (about 200 ml). In practice, V_A is about 94 ± 7% of TLC, 0.1–0.6 L less in absolute terms (Roberts et al., 1990). In airflow obstruction the single breath VA may be considerably less than the true TLC measured by multi-breath gas dilution or body plethysmography (Roberts et al., 1990).

Causes of a low Kco

The common causes of a low Kco are well known – particularly emphysema and diffuse alveolar–capillary damage associated with connective tissue/ autoimmune disease (see Table 1). In the earlier section 'Physiology of DL.co', we emphasized that most of the resistance to CO uptake lay within the microvasculature. Consequently, in lung disease, loss or destruction of the pulmonary capillary bed is a much more important mechanism for reducing Kco than thickening or inflammatory

Table 1 Some of the commoner causes of a Kco which is lower or higher than the reference value (adapted from Hughes & Pride, 2001).

change in the extravascular tissues. In the Churg–Strauss syndrome and in bronchiolitis (Table 1) a low Kco suggests vasculitis (the pulmonary arterioles and bronchioles share a common connective tissue sheath). In addition, a physiological cause of a low Kco is a reduced haemoglobin level, and it is customary to make a correction for this (American Thoracic Society, 1995; Cotes *et al.*, 1972, 1993).

Causes of a high Kco

This is a more difficult concept to grasp. There are physiological reasons for a high Kco (in terms of percentage predicted for a normal TLC): first, incomplete alveolar expansion (but without alveolar disruption) in which the lungs are not inflated to the level of the predicted TLC, secondly, an increase in pulmonary blood flow per unit lung volume. Kco, in terms of the Roughton–Forster equation, consists of two conductances, DM/VA and $\theta Vc/VA$ [θ may be ignored if PAO₂ and (Hb) are normal].

In the case of incomplete alveolar expansion in an otherwise normal lung, T_L/V_A or Kco increases linearly so that at FRC (~50% of V_A at full inflation, i.e. TLC) Kco is > 150% of Kco at TLC (Stam et al., 1994; Hughes & Pride, 2001. This Kco rise is caused mostly by an increase in the Vc/ V_A term (Vc does not change and V_A falls), while the DM/ V_A ratio remains constant or falls slightly (Stam et al., 1991). Clinically, a rise in Kco (up to 150% of that predicted for a normal TLC) will occur in neuromuscular, pleural or chest wall disease if TLC is reduced. Secondary factors, such as atelectasis or parenchymal disease, may limit the expected rise of Kco.

Low Kco	High Kco
Diffuse alveolar-capillary damage	Loss of units (discrete)
Pulmonary fibrosis	Pneumonectomy
Connective tissue/ autoimmune diseases	Local destruction / infiltrates
Sarcoidosis, asbestosis, bleomycin	
Pulmonary hypertension associated	Incomplete alveolar expansion:
Vasculitis	Pleural disease
Thromboembolic	Neuromuscular
Congestive heart failure / mitral stenosis	Chest wall deformity
Pulmonary oedema	Poor technique
Intrapulmonary shunting	Alveolar haemorrhage
Pulmonary arteriovenous malformations (PAVMs)	Anti-GBM disease
Hepatopulmonary syndrome (HPS)	Pulmonary vasculitis
	Wegener's granulomatosis
	SLE
	Idiopathic haemosiderosis
Airflow obstruction	Increased pulmonary blood flow
Emphysema	ASD
Churg–Strauss syndrome	Asthma
Bronchiolitis	
Low $\theta CO/V_{A}^{a}$	High $\theta CO/V_{A}^{a}$
Anaemia	Polycythaemia rubra vera
	Secondary polycythaemia

GBM, glomerular basement membrane; SLE, systemic lupus erythematosus; ASD, atrial septal defect.

^a Corrections can be made for an abnormal haemoglobin level.

Table 2 Different mechanisms reducing single breath VA in respiratory disease (adapted from Hughes & Pride, 2001).

Restrictive Disease with a small 1	LC and normal VA/TLC ratio	0	Obstructive Disease with normal or increased TLC
I	Ш	Ш	IV
Lack of lung expansion: lung structure normal	Loss of units: remaining lung structure normal	Diffuse alveolar damage	Sampled VA < TLC due to incomplete mixing during breath-holding
Examples			
Acute inspiratory muscle weakness. Chest wall disease and pleural disease.	Pneumonectomy. Local alveolar infiltrate, collapse, consolidation or local destruction	Fibrosing alveolitis. Pulmonary oedema, congestive heart failure, mitral stenosis, bleomycin lung, Wegener's granulomatosis.	Incomplete mixing may be associated with alveolar destruction (emphysema), space-occupying lesions (bullae) or normal alveolar structure (asthma)

An increase in pulmonary blood volume also increases Kco, because the Vc/VA ratio rises (as does the DM/VA ratio). Apart from an increase in pulmonary venous pressure (as in mitral stenosis or congestive heart failure) (Puri et al., 1995), the cause of the increase in Vc is an increase in cardiac output. On exercise Kco (and TL,CO) increase by about 20% per 5 l min⁻¹ increase in cardiac output from its resting value (Hsia et al., 1995). At rest, pulmonary blood flow may increase as a result of left to right shunts, or blood flow may become more 'homogeneous', as in asthma (Collard et al., 1994), representing an 'effective' blood flow increase. A much commoner situation in lung disease is where blood flow is diverted from diseased lung to normal lung, whose blood flow per unit volume increases. A clear-cut example is pneumonectomy, where blood flow per unit volume at rest doubles in the remaining lung, and Kco increases. Corris et al. (1987), in a study of 28 patients before and after pneumonectomy, found that the post-pneumonectomy Kco was 110-131% predicted. Hughes & Pride (2001) have referred to this situation as loss of alveolar units (discrete) where discrete means that some normal (unaffected) lung units are present. There may be many causes (see Table 1 and Table 2 [II]).

A low Kco will usually be associated with a low TL.CO, as it is unusual for VA to exceed its predicted normal value. A high Kco may be associated with a high, normal or a low TL.CO, depending (a) on the level of Kco, and (b) whether VA is normal or reduced. In alveolar haemorrhage, Kco is only elevated when active bleeding is taking place (Ewan et al., 1976), but the Hb-corrected Kco may be sufficiently high to raise TL.CO, although VA may be somewhat reduced.

Causes of a low VA

There are two causes of a low V_A ; restrictive lung disease when absolute lung volumes are small (Table 2; I–III), but the VA/TLC ratio is normal, and obstructive lung disease (Table 2, IV) where TLC is usually normal or increased, but the sampled V_A is <TLC due to incomplete mixing during breath holding. Restrictive lung disease can be subdivided further (see Table 2) into (I) lack

of alveolar expansion, (II) loss of alveolar units, discrete, and (III) diffuse alveolar-capillary damage.

Causes of a low TL,co

The same reduction in TLCO (say to 60% predicted) can be produced by several combinations of KCO and VA. Hughes & Pride (2001) have shown that, in restrictive lung disease, a TLCO of 60% predicted could be associated with (a) acute neuromuscular disease, (b) alveolar haemorrhage, (c) lung resection or collapse, (d) diffuse alveolar damage (connective tissue disease) or (e) pulmonary vascular pathology, depending on the precise combination of KCO and VA. Therefore, inspection of the components of TLCO (KCO and VA) is essential if a reasonable interpretation of the TLCO test is to be made.

Clinical examples

Table 3 lists examples of a low TLCO in some pulmonary diseases. In inspiratory muscle weakness (Hart et al., 2002), KCO is 130% predicted (for a normal TLC), but at this low VA (50% predicted) the KCO for a normal subject would be >150% predicted (Hughes & Pride, 2001). As previously mentioned, parenchymal lung damage or atelectasis limit the rise in KCO in 'extrapulmonary' restriction.

The Kco for a 50% reduction in VA due to loss of lung units is about 115% predicted (Hughes & Pride, 2001), and this is what occurs post-pneumonectomy (Corris *et al.*, 1987). The situations in Table 3 where Kco is normal (sarcoidosis, bronchiectasis) but VA is reduced (and there is no airflow obstruction as a cause) are compatible with loss of lung units due to underlying disease with sparing of the remainder of the lung.

Where Kco is moderately impaired (84-85%) in CHF and fibrosing alveolitis, there is presumably a spectrum of lung damage – normal units with a Kco in the 100–110% range and diseased units with a low Kco (<80%). In Table 3, note four situations (post-pneumonectomy, fibrosing alveolitis, primary PHT and emphysema) where TL_{CO} is essentially the same (54– 58% predicted) but inspection of the VA and Kco patterns (and, in the case of emphysema, the FEV₁/VC ratio) reveals different

Diagnosis	Source	FEV ₁ /VC (actual)	Vc %	Va %	Kco%	Тг,со%	Comment
High/normal Kco with low V _A							
Inspiratory muscle weakness	Hart (2002)	0-79	50	55	130	66	Lack of alveolar expansion (see text)
Pneumonectomy	Corris (1987)	NA	82*	77*	88 *	78*	*Pre **Post
		NA	53**	51**	111^{**}	58**	Loss of lung units, discrete
Sarcoidosis with infiltrates	Hughes (1999)	0.8	83	75	105	76	Loss of lung units predominantly
Bronchiectasis§	Perez (1998)	0.75#	96	82#	66	82	Loss of lung units predominantly
Low Kco with low/normal V _A							
CHF (NYHA III)	Puri (1995)	0.71	76	75	85	72	Loss of units; some microvascular damage
Fibrosing alveolitis	Hughes (1999)	0.78	73	66	84	54†	Loss of units; some alveolar–capillary damage
Primary PHT	Hughes (1999)	6-0	91	100	58	58†	Diffuse microvascular damage
Emphysema	Gevenois (1996)	0.55	84	74	49	54†	Diffuse alveolar–capillary damage

Ξ.

PHT, pulmonary hypertension; CHF, congestive heart failure; NYHA III, New York Heart Association, grade

four instances, but different combinations of VA and

in these

TL,CO

Note similar

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pathophysiological mechanisms in each case see Hughes (1999) for more examples.

TL, co, exercise and arterial hypoxaemia

The TL,CO is rarely measured on exercise except in research protocols. In normal subjects, TL,CO increases in proportion to the workload; there is no evidence of a plateau (Hsia et al., 1995). At near maximal exercise, TL,CO is 50% higher than resting values in men (25% higher in women) (Hsia et al., 1995). In patients with lung disease, the fractional (percentage) increase in TL,CO for a given level of exercise is similar to that in normal subjects (reviewed in Hughes, 1991), although exercise levels are quite low ($\dot{V}O_2 \le 1.0 \text{ Lmin}^{-1}$). When resting TL,co is <60% predicted, worsening of arterial hypoxaemia on exercise is seen almost invariably. Patients with interstitial lung disease and diffuse alveolar-capillary damage have been the subjects of most studies. It is not easy to differentiate diffusion limitation from $\dot{V}_{\rm A}/\dot{\rm Q}$ mismatch as causes of the exercise-induced hypoxaemia - a problem first studied in the 1940s (Baldwin et al., 1949).

In interstitial lung disease patients with a low TL,CO, diffusion limitation, in terms of the contribution of the PA-Pc' (alveolar to end-capillary) gradient to the overall PA-Pa (alveolar to arterial) oxygen gradient, is about 10% at rest, 20-30% on light exercise and more than 50% at a $\dot{V}O_2$ of 1.0 L min⁻¹ (Hughes, 1991). The reason for the widening PA-Pc' gradient (failure of capillary PO₂ to equilibrate with alveolar PO₂ before red cells leave the alveolus) is a low diffusion-perfusion ratio (~TL,CO/ \dot{Q}) within gas exchanging units, or for the lung overall if the disease process is diffuse. In interstitial lung disease, the TL/\dot{Q} ratio at rest is generally above the critical threshold for diffusion limitation, but on exercise, the increase in TL,CO, starting from a low base, is insufficient to match the increase in blood flow, so the TL/ \dot{Q} ratio falls and a PA-Pc' gradient emerges. On the other hand, in patients with airflow obstruction (emphysema, for example), any change in PaO₂ on exercise will reflect ventilatory limitation and local $\dot{V}_{\rm A}/\dot{Q}$ mismatching more than any decline in T_L/\dot{Q} ratios.

Conclusion

Recent physiological evidence, using the Roughton–Forster analysis, and morphometric measurements on lungs postmortem, suggest that most of the resistance to CO transfer from alveolar gas to pulmonary capillary blood may lie in the red cell itself. The clinical implication is that TL,co and Kco are focused primarily on the pulmonary capillary bed. Although non-specific diffuse alveolar damage, as in interstitial fibrosis, or alveolar destruction, as in emphysema, will compromise the microvasculature and reduce TL,co and Kco, pathology specific to the pulmonary circulation, without diffuse alveolar damage, such as vasculitis, raised pulmonary venous pressure or microvascular dilatation (as in PAVMs and HPS) also reduces TL,co and Kco. As simple monitors of the integrity of the pulmonary microcirculation, TL,CO and KCO are uniquely valuable and important clinical tests.

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